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Supplementary Appendix S1: Short protocol for the isolation of high molecular weight genomic DNA (96-well format).

1. Grind samples with stainless steel beads in 1.2 ml collection microtubes (Qiagen) in a Tissue Lyzer II mixer mill to a fine powder.
2. Add 400 µl CTAB lysis buffer (heated to 65°C) & 1 µl RNase A (optional), mix vigorously until all powder has been dissolved.
3. Spin at 6000 g for 10 minutes at 15–20°C.
4. Pipet the supernatant into fresh collection microtubes. Then add 600 µl binding buffer.
5. Mix well. Pipet binding buffer mix (1000 µl) on the glass fibre membrane plate residing on a 2.2 ml deep-well plate.
6. Spin for 3 minutes*, discard flow-through.
7. Wash the membrane by adding 500 µl 70% ethanol to each well. Spin for 3 minutes*, then discard the flow-through. Repeat wash and discard the flow-through. Spin for 15 minutes* to remove residual ethanol from the membranes.
8. Place the glass fibre membrane plate on a clean deep-well plate (autoclaved). Elute genomic DNA with 110 µl AE buffer preheated to 65°C, incubate 1 minute, spin 1 min*. Repeat elution to gain a total of 220 µl of DNA.

* Spinning at 3800 g and 15–20°C. If liquid has not passed through membrane, increase centrifugation time.